Anti-Jo-1

ORG 513

96 Tests

Immunometric Enzyme Immunoassay for the quantitative determination of anti-Jo-1 Antibodies

Instruction for use
WARNINGS AND PRECAUTIONS

All reagents of this test kit are strictly intended for in vitro use only. Please adhere strictly to the sequence of pipetting steps provided in this protocol. Observe the guidelines for performing quality control in medical laboratories by assaying controls and/or pooled sera.

All reagents should be stored refrigerated at 2 - 8 °C in their original container. Do not interchange kit components from different lots. The expiration dates stated on the labels of the shipping container and all vials have to be observed. Do not use kit components beyond their expiration dates.

Allow all kit components and specimen to reach room temperature prior to use and mix well. During handling of all kit reagents, controls and serum samples observe the existing legal regulations. The following precautions should be taken handling potentially infectious materials:

- do not eat, drink or smoke in areas where specimens or kit reagents are handled
- do not pipette by mouth
- wear disposable gloves while handling specimens or kit reagents and wash hands thoroughly afterwards.

The test kit contains components of human origin which, when tested by FDA-licensed methods, were found negative for hepatitis B surface antigen and for HIV antibody. No known test can guarantee, however, that products derived from human blood will not be infectious. Handle, therefore, all reagents and human blood derivatives, like plasma or serum samples, as if capable of transmitting infection.

Avoid contact with the TMB (3,3',5,5'-Tetramethyl-benzidine). If TMB comes into contact with skin wash thoroughly with water and soap.

The stop solution contains hydrochloric acid. If it comes into contact with skin, wash thoroughly with water and seek medical attention.

Avoid contact between the buffered Peroxide Solution and easily oxidized materials; extreme temperatures may initiate spontaneous combustion.

MATERIALS SUPPLIED

Package size 96 determ.

- divisible microplate consisting of 12 modules of 8 wells, .................... 1
- coated with highly purified Jo-1
- Anti-Jo-1 calibrators in a PBS/BSA matrix ........................................ 6 vials, 1.5 ml each containing respectively: 0; 12.5; 25; 50; 100 and 200 U/ml
- Anti-Jo-1 controls in a serum/buffer matrix ................................. 2 vials, 1.5 ml each (positive and negative), for the respective concentrations see the enclosed package insert
- sample buffer, yellow, concentrate ............................................. 1 vial, 20 ml
- enzyme conjugate solution, (light red) containing polyclonal .......... 1 vial, 15 ml rabbit anti-h-IgG-IgG; labelled with horseradish peroxidase
- TMB substrate solution ............................................................... 1 vial, 15 ml
- stop solution (1 M hydrochloric acid) ........................................ 1 vial, 15 ml
- buffered wash solution, concentrate ........................................... 1 vial, 20 ml
CONTROLS

A set of two controls is provided with the kit.

TECHNICAL DATA

Sample material: serum or plasma
Required sample volume: 10 µl of sample to be diluted 1:100 with sample buffer
100 µl prediluted sample per single determination
Total incubation time: 60 minutes at room temperature (20 - 28 °C)
Calibration range: 12.5 – 200 U/ml
Sensitivity: 1 U/ml
Storage: refrigerated at 2 - 8 °C
Shelf life: 12 months after manufacturing or until the expiration date printed on the labels
Package size: 96 tests

PRINCIPLE OF THE PROCEDURE

Anti-Jo-1 is an indirect solid phase enzyme immunometric assay (ELISA). It is designed for the quantitative measurement of IgG class autoantibodies directed against the extractable nuclear antigens Jo-1. The microplate is coated with Jo-1.

The microplate can be divided into 12 modules of 8 wells each or can be used completely for 96 determinations. Each well can be separated from the module ("break-away"). The wells of the microplate are stained with a specific color for each ENA antigen:

<table>
<thead>
<tr>
<th>assay</th>
<th>anti-SS-A</th>
<th>anti-SS-B</th>
<th>anti-Sm</th>
<th>anti-RNP/Sm</th>
<th>anti-Scl 70</th>
<th>anti-Jo-1</th>
</tr>
</thead>
<tbody>
<tr>
<td>color code</td>
<td>red</td>
<td>blue</td>
<td>yellow</td>
<td>green</td>
<td>pink</td>
<td>black</td>
</tr>
</tbody>
</table>

The binding of present autoantibodies, as well as the formation of the sandwich complexes and enzymatic color reaction take place during three different reaction phases:

**Phase 1:**
Calibrators, controls and prediluted patient samples are pipetted into the wells of the microplate. Any present antibodies bind to the inner surface of the wells. After a 30 minutes incubation the microplate is washed with wash solution for removing non-reactive serum components.

**Phase 2:**
An anti-human-IgG horseradish peroxidase conjugate solution is pipetted into the wells of the microplate to recognize the autoantibodies bound to the immobilized antigens. After a 15 minutes incubation any excessive enzyme conjugate, which is not specifically bound is washed away with wash solution.

**Phase 3:**
A chromogenic substrate solution containing TMB (3,3',5,5'-Tetramethyl-benzidine) is dispensed into the wells. During 15 minutes of incubation the color of the solutions change into blue. Color development is stopped by adding 1 M hydrochloric acid as stop solution. The solutions color change into yellow. The amount of colour is directly proportional to the concentration of IgG present in the original sample.
To read the optical density a microplate reader with a 450 nm filter is required. Bi-chromatic measurement with a 600-690 nm reference is recommended. The optical density for each calibrator may be graphically plotted against the concentration of IgG and unknowns extrapolated from the curve.

**CLINICAL RELEVANCE**

Rheumatoid autoimmune diseases are often associated with the occurrence of autoantibodies against several nuclear or cytoplasmatic antigens. These so-called anti nuclear antigens (ANA) can be divided into three groups:

1. true anti nuclear antigens (ANA): dsDNA, ssDNA, histones, nucleolic RNA and DNP
2. extractable nuclears antigens: Sm (Smith), n-RNP, Scl 70 and PM-1
3. cytoplasmatic antigens: SS-A (Ro)*, SS-B (La)* and Jo-1

SS-A (Ro) and SS-B (La) are co-localized in cytoplasm and nucleus

In patients with Sjögren-Syndrome antibodies against SS-A and SS-B often occur in combination. Due to the strong association of SS-A and SS-B antibodies to the HLA-DR3 and DR2 phenotypes a genetic predisposition is suspected. The anti SS-A protein passes the placenta and may cause the development of SLE in neonates.

Immunoreactive proteins may occur in various combinations and bind also to 'host proteins' of viral origin. They induce synthesis of polyclonal autoantibodies, of the IgG, IgM and IgA class. Especially for mixed connective tissue diseases a relation to viral infections by EBV (Eppstein-Barr-Virus) is indicated.

Each class of immunoglobulins causes a specific immunofluorescent pattern. Basically immunofluorescence titers correlate with the quantitation of IgG antibodies but the concentrations may vary considerably within each titer. Quantitation of IgG class antibodies extensively correlates with the diseases' activity. This makes it superior to immuno-fluorescence using Hep2 cells. The IF with Crithidia lucilliae sometimes results in deviating values.

Today the best investigated immunoreactive antigens are double-stranded DNA (dsDNA), single stranded DNA (ssDNA), Sm (Smith), sn-RNP (small nuclear ribonucleoprotein particles), the complex RNP/Sm which is stabilized by ribonucleic acid as well as SS-A (Ro) and SS-B (La). The antigen Scl 70, a 70 kD molecular weight protein is associated with scleroderma.

In rheumatoid autoimmune diseases various profiles of autoantibodies to these antigens can be detected. In a high incidence they are related to active and inactive systemic Lupus erythematosides, mixed connective tissue diseases (Sharp Syndrome), rheumatoid arthritis, Sjögren-Syndrome, Scleroderma, photosensitive dermatitis and drug-induced lupus.

In Lupus patients typically anti-dsDNA antibodies can be detected. Patients without these antibodies very often show anti-ssDNA antibodies and anti-SS-A and anti-SS-B are present. A strong correlation between antibody concentration and severity of the disease has been observed with higher antibody concentrations in active phases of the disease. Thus quantitation is more informative compared to simple titering by immunofluorescence.

Measurement of anti-ssDNA provides additional information regarding antibody specificity and activity. Except in chronic inflammatory processes anti-ssDNA antibodies are not found in healthy subjects.

Most of these parameters are not specific for just one disease but they occur in various combinations. The pattern of different antibody combinations and their concentration together with the whole clinical picture of the patient are helpful diagnostic tools in the assessment of rheumatoid autoimmune diseases.

The following graph gives brief information on the complexity of autoimmune diseases, occuring antibodies. It is not disigned as a diagnostic schedule or program for ongoing diagnostic profiles.


<table>
<thead>
<tr>
<th>Diseases (values in %)</th>
<th>dsDNA</th>
<th>ssDNA</th>
<th>Histone</th>
<th>SS-A (Ro)</th>
<th>SS-B (La)</th>
<th>Sm</th>
<th>RNP/Sm</th>
<th>Sm/Scl 70</th>
<th>Jo-1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Systemic Lupus erythematoses (SLE)</td>
<td>&gt; 90</td>
<td>&gt; 90</td>
<td>30-50</td>
<td>10-30</td>
<td>30-50</td>
<td>10-30</td>
<td>10-30</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Drug induced Lupus (LE)</td>
<td>30-50</td>
<td>50-90</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sharp-Syndrome / Mixed connective tissue diseases</td>
<td>10-30</td>
<td>10-30</td>
<td>&gt; 90</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rheumatoid Arthritis</td>
<td>30-50</td>
<td>30-50</td>
<td>10-30</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sjögren’s syndrome</td>
<td>10-30</td>
<td>10-30</td>
<td>&gt; 90</td>
<td>&gt; 90</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Scleroderma</td>
<td>10-30</td>
<td>10-30</td>
<td>10-30</td>
<td>&gt; 90</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Photosensitive Dermatitis Dermatomyositis</td>
<td>10-30</td>
<td>10-30</td>
<td>50-90</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>


NORMAl VALUES

In a normal range study with serum samples from healthy blood donors the following ranges have been established with the Anti-Jo-1 test:

\[
\text{Anti-Jo-1} [\text{U/ml}]
\]
- normal: < 15
- borderline: 15 - 25
- elevated: > 25

Positive results should be verified concerning the entire clinical status of the patient. Also every decision for therapy should be taken individually. It is recommended that each laboratory establishes its own normal and pathological ranges of serum Anti-Jo-1 antibodies. The above reference ranges should be regarded as guidelines only.

SPECIFICITY

The microplate is coated with Jo-1 highly purified by affinity chromatography. The Anti-Jo-1 test kit is specific only for autoantibodies directed to Jo-1. No crossreactivities to the other ENA-antigens have been observed.
CALIBRATION

The assay system is calibrated against the internationally recognized reference sera from CDC, Atlanta USA, since no other international standards are available.

REFERENCES

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MATERIALS REQUIRED

Equipment
- Microplate reader capable for endpoint measurements at 450 nm
- Vortex mixer
- Pipets for 10 µl, 100 µl and 1000 µl

Preparation of reagents
- distilled water
- graduated cylinder for 100 and 1000 ml
- plastic container for storage of the wash solution

Optional
- Multi-Chanel Dispenser
- or repeatable pipet for 100 µl
- data reduction software

SPECIMEN COLLECTION AND PREPARATION

For determination of Anti-Jo-1 antibodies serum or plasma are the preferred sample matrixes. All serum and plasma samples are pre-diluted 1 : 100 with sample buffer. Therefore 10 µl of sample may be diluted with 1000 µl of sample buffer. The patients need not to be fasting, and no special preparations are necessary. Collect blood by venipuncture into vacutainers and separate serum or plasma from the cells by centrifugation after clot formation. Samples may be stored refrigerated at 2 - 8 °C for at least 5 days. For longer storage of up to six months samples should be stored frozen at -20 °C. To avoid repeated thawing and freezing the samples should be aliquoted. Neither Bilirubin nor Hemolysis have significant effect on the procedure.

PREPARATION AND STORAGE OF REAGENTS

All components of this test kit are supplied in a liquid format and ready to use, except the sample buffer and wash buffer. When stored refrigerated at 2 - 8 °C the components are stable for at least 30 days after opening or until the expiration date printed on the labels. Remaining modules of the microplate should be stored refrigerated at 2 - 8 °C protected from moisture; store together with desiccant and carefully sealed in the plastic bag.

Preparation of sample buffer
Dilute the contents of each vial of the sample buffer concentrate (5x) with distilled water to a final volume of 100 ml prior to use. Store refrigerated: stable at 2 - 8 °C for at least 30 days after preparation or until the expiration date printed on the label.

Preparation of buffered wash solution
Dilute the contents of each vial of the buffered wash solution concentrate (50x) with distilled water to a final volume of 1000 ml prior to use. Store refrigerated: stable at 2 - 8 °C for at least 30 days after preparation or until the expiration date printed on the label.
NOTES ON TECHNIQUE

Control sera or pools should routinely be assayed as unknowns to check performance of the reagents and the assay.

For all controls, the respective concentrations are provided on the labels of each vial. Using these concentrations a calibration curve may be calculated to read off the patient results semi-quantitatively.

Pipetting and Sample Handling
Use a disposable-tip micropipette to dispense sera and plasma samples. Pipet directly to the bottom of the wells. To avoid carryover contamination change the tip between samples.

Patient samples expected to contain high concentrations should be additionally diluted with sample buffer before. Additional dilutions must be considered during calculation.

ASSAY CHARACTERISTICS

Sensitivity
The lower detection limit for the Anti-Jo-1 test was determined at 1 U/ml.

Parallelism
In dilution experiments sera with high antibody concentrations were diluted with sample buffer and assayed in the Anti-Jo-1 kit. The assay showed linearity over the full measuring range.

Precision
Statistics were calculated for each of three samples from the results of 24 determinations in a single run for Intra-Assay precision and the run-to-run precision was calculated from the results of 5 different runs with 6 determinations each:

<table>
<thead>
<tr>
<th>Intra-Assay</th>
<th>Inter-Assay</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample No</td>
<td>Mean (U/ml)</td>
</tr>
<tr>
<td>1</td>
<td>32.2</td>
</tr>
<tr>
<td>2</td>
<td>73.2</td>
</tr>
<tr>
<td>3</td>
<td>134.0</td>
</tr>
</tbody>
</table>
IMMUNOASSAY PROCEDURE

Do not interchange components of different lots.
All components should be at room temperature before use.
Dilute all patient samples 1:100 with sample buffer before assay. Therefore combine 10 µl of
sample with 1000 µl of sample buffer in a polystyrene tube. Mix well. Calibrators and controls are
ready to use and need not to be diluted.
1. Prepare a sufficient number of microplate modules to accommodate calibrators, controls and
   prediluted patient samples in duplicates.

<table>
<thead>
<tr>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>SA</td>
<td>SE</td>
<td>P1</td>
<td>P5</td>
<td></td>
</tr>
<tr>
<td>B</td>
<td>SA</td>
<td>SE</td>
<td>P1</td>
<td>P5</td>
<td></td>
</tr>
<tr>
<td>C</td>
<td>SB</td>
<td>SF</td>
<td>P2</td>
<td>P..</td>
<td></td>
</tr>
<tr>
<td>D</td>
<td>SB</td>
<td>SF</td>
<td>P2</td>
<td>P..</td>
<td></td>
</tr>
<tr>
<td>E</td>
<td>SC</td>
<td>C1</td>
<td>P3</td>
<td></td>
<td></td>
</tr>
<tr>
<td>F</td>
<td>SC</td>
<td>C1</td>
<td>P3</td>
<td></td>
<td></td>
</tr>
<tr>
<td>G</td>
<td>SD</td>
<td>C2</td>
<td>P4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>H</td>
<td>SD</td>
<td>C2</td>
<td>P4</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

   SA - SF: standards A to F
   P1, P2... patient sample 1, 2...
   C1: positive control
   C2: negative control

2. Pipet 100 µl of calibrators, controls and prediluted patient samples into the wells.
3. Incubate for 30 minutes at room temperature (20 - 28 °C).
4. Discard the contents of the microwells and wash 3 times with 300 µl of wash solution.
5. Dispense 100 µl of enzyme conjugate solution into each well.
6. Incubate for 15 minutes at room temperature.
7. Discard the contents of the microwells and wash 3 times with 300 µl of wash solution.
8. Dispense 100 µl of TMB substrate solution into each well.
9. Incubate for 15 minutes at room temperature protected from light.
10. Add 100 µl of stop solution to each well of the modules and leave untouched for 5 minutes.
11. Read the optical density at 450 nm and calculate the results. Bi-chromatic measurement with
    reference at 600-650 nm is recommended.

    The developed color is stable for at least 30 minutes.
    Read optical densities during this time.

CALCULATION OF RESULTS

For the Anti-Jo-1 test a 4-Parameter-Fit with lin-log coordinates for optical density and
concentration is recommended. Spline Approximation and log-log coordinates are also suitable.
Recommended Lin-Log Plot
First calculate the averaged optical densities for each calibrator well. Use lin-log graph paper and
plot the averaged optical density of each calibrator versus the concentration. Draw the best fitting
curve approximating the path of all calibrator points. The calibrator points may also be connected
with straight line segments. The concentration of unknowns may then be estimated from the
 calibration curve by interpolation.
CALCULATION EXAMPLE

The figures below show typical results for Anti-Jo-1 test. These data are intended for illustration only and should not be used to calculate results from another run.

<table>
<thead>
<tr>
<th>No</th>
<th>Position</th>
<th>OD 1</th>
<th>OD 2</th>
<th>Mean</th>
<th>Conc. 1</th>
<th>Conc. 2</th>
<th>Mean decl.</th>
<th>Conc. CV %</th>
</tr>
</thead>
<tbody>
<tr>
<td>STA</td>
<td>A 1 / B 1</td>
<td>0.032</td>
<td>0.033</td>
<td>0.033</td>
<td>0.1</td>
<td>0.1</td>
<td>0.1</td>
<td>0</td>
</tr>
<tr>
<td>STB</td>
<td>C 1 / D 1</td>
<td>0.313</td>
<td>0.305</td>
<td>0.309</td>
<td>12</td>
<td>12</td>
<td>12</td>
<td>12</td>
</tr>
<tr>
<td>STC</td>
<td>E 1 / F 1</td>
<td>0.569</td>
<td>0.556</td>
<td>0.563</td>
<td>25</td>
<td>23</td>
<td>25</td>
<td>25</td>
</tr>
<tr>
<td>STD</td>
<td>G 1 / H 1</td>
<td>0.948</td>
<td>0.940</td>
<td>0.944</td>
<td>51</td>
<td>50</td>
<td>51</td>
<td>50</td>
</tr>
<tr>
<td>STE</td>
<td>A 2 / B 2</td>
<td>1.422</td>
<td>1.423</td>
<td>1.423</td>
<td>98</td>
<td>99</td>
<td>98</td>
<td>100</td>
</tr>
<tr>
<td>STF</td>
<td>C 2 / D 2</td>
<td>1.989</td>
<td>1.973</td>
<td>1.981</td>
<td>204</td>
<td>199</td>
<td>201</td>
<td>200</td>
</tr>
</tbody>
</table>

INCUBATION SCHEME

1. **Pipet 100 µl** calibrator, control or diluted patient sample
   - **Incubate for 30 minutes** at room temperature
   - **Discard the contents of the wells and wash 3 times with 300 µl** wash solution

2. **Pipet 100 µl** enzyme conjugate
   - **Incubate for 15 minutes** at room temperature
   - **Discard the contents of the wells and wash 3 times with 300 µl** wash solution

3. **Pipet 100 µl** substrate solution
   - **Incubate for 15 minutes** at room temperature

4. **Add 100 µl** stop solution
   - **Leave untouched for 5 minutes**
   - **Read at 450 nm**